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Stereoselective total synthesis of arenastatin A, a spongean cytotoxic depsipeptide

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Abstract—A highly stereoselective total synthesis of arenastatin A, an extremely potent cytotoxic cyclic depsipeptide from marine sponge, was developed. The desired 7,8- β -epoxide in arenastatin A was constructed by asymmetric sulfur ylide-mediated epoxidation in good yield and highly stereoselective manner. © 2007 Elsevier Ltd. All rights reserved.

In the course of our search for bioactive substances from marine organisms, we isolated and characterized arenastatin A (1), a cyclic depsipeptide having an extremely potent antiproliferative activity against KB cells (IC₅₀ 5 pg/mL), from the Okinawan marine sponge of *Dysidea arenaria*.¹ Thereafter, we achieved the first total synthesis of $1,^2$ and many synthetic studies toward 1 and its analogues have been reported so far, because of its synthetically attractive structure and potent biological activity (Fig. 1).³

Arenastatin A (1) has a $7R, 8R-\beta$ -epoxy moiety on its molecule. Some structure-activity relationship (SAR) studies have revealed that the β -epoxy moiety plays critical role for the potent cytotoxic activity of 1, and the α -epoxy isomer has no biological activity. However,



Figure 1. Chemical structure of arenastatin A (1).

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the epoxy function in 1 is labile under acidic condition,

so it should be introduced at the final stage of the syn-

thesis. The most reliable method for the stereoselective

construction of the 7,8- β -epoxide would be the epoxidation of the 7,8-E-olefin of **2** with dimethyldioxirane

developed by our group (Scheme 1).^{2a,4} However, the

stereoselective ratio is $\beta:\alpha = 2.2:1$, and reversed-phase

HPLC separation should be needed to isolate pure prod-



Scheme 1.

Keywords: Arenastatin A; Depsipeptide; Total synthesis; Asymmetric epoxidation.

Shi epoxidation of the substrate having cyclic depsipeptide structure resulted in very low conversion, because of steric hindrance. Thus, another method for the construction of the 7,8- β -epoxide in a highly stereoselective manner is strongly needed from the viewpoint of synthetic utility and atom economy. Here, we present our new synthetic procedure for the stereoselective total synthesis of arenastatin A (1) by using asymmetric sulfur ylidemediated epoxidation.⁵

The synthetic strategy was shown in Figure 2. An aryl moiety and a 7R,8R-epoxide of 1 could be introduced in one operation by the Corey–Chaycovsky reaction between a chiral benzyl sulfur ylide and an aldehyde 3 having cyclic depsipeptide structure. Compound 3 could be synthesized through the similar manner as our previous total synthesis of $1,^2$ that is, the successive condensation of the four segments A (6), B (5), C (7), and D (8).

Cyclic peptide **3** was synthesized as follows (Schemes 2, 3). First of all, segment A (**6**) was prepared with a known method.⁶ Thus, Lewis acid-promoted allylation of aldehyde **9** gave a homoallylic alcohol **10**, as a mix-

ture of the diastereomers in a 13/1 ratio. Removal of *p*-methoxybenzyl (MPM) group of **10** and subsequent protection of the primary hydroxyl group of **11** by *tert*-butyldiphenylsilyl (TBDPS) group afforded a desired segment A (**6**).

Segment A (6) was coupled with segment C (7), prepared from L-leucic acid,² to give segment AC (12) in 87%yield. Oxidative cleavage of the terminal olefin of 12 smoothly proceeded to give an aldehyde 4, and subsequent Horner-Emmons reaction with segment B (5) afforded an α,β -unsaturated amide 13. It revealed that the choice of base was important in this reaction. The results are summarized in Table 1. Thus, the use of NaH as a base (entry 1) resulted in disappointing yield of the desired product, and considerable amount of α,β -unsaturated aldehyde 14 was obtained as a byproduct. On the other hand, the Masamune-Roush condition⁷ using DBU and LiCl (entry 2), which was developed for base-sensitive aldehydes, afforded 14 as a sole product. Among tested, the use of activated $Ba(OH)_2^8$ (entry 6) gave the best result to afford the coupling product 13 in 60% yield (three steps from 12).



Figure 2. Retrosynthetic analysis.



Scheme 2. Reagents and conditions: (a) allyltributylstannane, SnCl₄, CH₂Cl₂, -78 °C, 75%, 13:1 dr; (b) HCl/MeOH, reflux, 80%; (c) TBDPSCl, Et₃N, DMAP, CH₂Cl₂, 95%; (d) 7, EDCI·HCl, DMAP, CH₂Cl₂, 87%; (e) OsO₄, NMO, THF/pH 7.0 buffer; (f) NaIO₄, THF/pH 7.0 buffer; (g) 5, Ba(OH)₂, THF/H₂O (40:1), 60% (three steps).



Scheme 3. Reagents and conditions: (a) BF_3 ·Et₂O, PhSH, CH_2Cl_2 , -45 °C, 71%; (b) 8, EDCI·HCl, DMAP, CH_2Cl_2 , 98%; (c) TFA, CH_2Cl_2 , 0 °C; (d) DPPA, NaHCO₃, DMF, 61% (two steps); (e) HF-pyridine, THF, 84%; (f) Dess-Martin periodinane, CH_2Cl_2 .

 Table 1. Conditions of Horner–Emmons reaction between 4 and 5

| Entry | Base | Yield ^a (%) |
|-------|------------------------|------------------------|
| 1 | NaH | 33 |
| 2 | LiCl, DBU | Decomp. |
| 3 | t-BuOK | Decomp. |
| 4 | n-BuLi, DMSO | 55 |
| 5 | $Ba(OH)_2$ (2.0 equiv) | 56 |
| 6 | $Ba(OH)_2$ (0.8 equiv) | 60 |

^a Isolated yield of **13** in three steps from **12**.

Removal of the MPM group of 13 using PhSH and $BF_3 \cdot Et_2O$ and subsequent condensation with segment D (8) using the conventional method^{2a} afforded 15 in good yield. Finally, removal of both (trimethylsilyl)ethyl group and *tert*-butoxycarbonyl (Boc) group by trifluoro-acetic acid (TFA) and subsequent intramolecular macrocyclization using diphenylphosphoryl azide (DPPA) furnished a cyclic depsipeptide 16. Removal of the TBDPS protecting group and Dess-Martin oxidation gave a desired aldehyde 3 (Scheme 3).

In the final stage of the synthesis, that is, the introduction of the aryl moiety and 7R,8R-epoxide, we used asymmetric Corey–Chaycovsky reaction mediated by D-camphor-derived chiral sulfide, which has been developed by Aggarwal's group.⁵ In order to optimize the reaction condition, the condensation between aldehyde **18** and sulfur ylide, which was generated by deprotonation from sulfonium salt **19** using phosphazene base P₂–Et, was examined (Scheme 4). In the case using



Scheme 4.

stoichiometric amount, α , β -unsaturated aldehyde **21** was obtained as a sole product, which was produced by β -elimination of the leucine moiety from **18**. The use of increased amount of the sulfur ylide tended to give a desired epoxide **20** together with an epoxide **22**, which was produced from **21** and sulfur ylide. The ¹H NMR spectrum and HPLC analysis of **20** revealed that epoxide **20** was obtained as almost single stereoisomer! The stereochemistry of **20** was ascertained by NOESY experiment. However, even in the optimized reaction condition, the desired epoxide **20** was obtained in low yield (~25%), and the β -elimination reaction could not be avoided.

Fortunately, the same reaction using cyclic peptide **3** as a substrate proceeded smoothly to give arenastatin A (**1**) in good yield (79%, two steps from **17**). The side product derived from the elimination of the ester moiety was not detected, probably because of the stability of the cyclic structure in **3**. The HPLC analysis of the reaction product revealed that the reaction proceeded with excellent stereoselectivity (β : $\alpha = \sim 100$:1, trans:cis = ~ 30 :1).^{2a,9} Recently, Sherman and co-workers¹⁰ reported that the stereoselective epoxidation of the 7,8-*E*-olefin in the cyclic precursor of cryptophycin 2 was succeeded by using cryptophycin CYP450 epoxidase, which was obtained by overexpression of the cyanobacterial gene in *Escherichia coli*.

In summary, we developed a convergent synthetic procedure of arenastatin A(1) by using asymmetric sulfur ylide-mediated epoxidation as a key step. This method provided a highly stereoselective construction of the $7R_{,8}R_{-\beta}$ -epoxide moiety in **1**.

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References and notes

- (a) Kobayashi, M.; Aoki, S.; Ohyabu, N.; Kurosu, M.; Wang, W.; Kitagawa, I. *Tetrahedron Lett.* **1994**, *35*, 7969– 7972; (b) Kobayashi, M.; Kurosu, M.; Ohyabu, N.; Wang, W.; Fujii, S.; Kitagawa, I. *Chem. Pharm. Bull.* **1994**, *42*, 2196–2198.
- (a) Kobayashi, M.; Kurosu, M.; Wang, W.; Kitagawa, I. *Chem. Pharm. Bull.* **1994**, *42*, 2394–2396; (b) Kobayashi, M.; Wang, W.; Ohyabu, N.; Kurosu, M.; Kitagawa, I. *Chem. Pharm. Bull.* **1995**, *43*, 1598–1600.

- (a) Eißler, S.; Stoncius, A.; Nahrwold, M.; Sewald, N. Synthesis 2006, 3743–3789; (b) Eggen, M.; Georg, G. I. Med. Res. Rev. 2002, 22, 85–101; (c) Tius, M. A. Tetrahedron 2002, 58, 4343–4367.
- Hoard, D. W.; Moher, E. D.; Martinelli, M. J.; Norman, B. H. Org. Lett. 2002, 4, 1813–1815.
- (a) Aggarwal, V. K.; Winn, C. L. Acc. Chem. Res. 2004, 37, 611–620; (b) Li, A.-H.; Dai, L.-X.; Aggarwal, V. K. Chem. Rev. 1997, 97, 2341–2372.
- (a) White, J. D.; Hong, J.; Robarge, L. A. J. Org. Chem. 1999, 64, 6206–6216; (b) Keck, G. E.; Park, M.; Krishnamurthy, D. J. Org. Chem. 1993, 58, 3787–3788.
- Blanchette, M. A.; Choy, W.; Davis, J. T.; Essenfeld, A. P.; Masamune, S.; Roush, W. R.; Sakai, T. *Tetrahedron Lett.* 1984, 25, 2183–2186.
- Paterson, I.; Yeung, K.-S.; Smaill, J. B. Synlett 1993, 774– 776.
- 9. The isomeric ratio of the epoxide was determined by reversed-phase HPLC. The stereochemistry of the *cis*-epimer was deduced by the coupling constant of the ¹H NMR signal of H-8 (δ 4.05, J = 4.5 Hz), which was characteristic for the *cis*-epoxide.
- Magarvey, N. A.; Beck, Z. Q.; Golakoti, T.; Ding, Y.; Huber, U.; Hemscheidt, T. K.; Abelson, D.; Moore, R. E.; Sherman, D. H. *ACS Chem. Biol.* **2006**, *1*, 766–779.